There is good evidence that telomere shortening acts as a biological clock in human fibroblasts, limiting the number of population doublings a culture can achieve. Oxidative stress also limits the growth potential of human cells, and recent data show that the effect of mild oxidative stress is mediated by a stress-related increased rate of telomere shortening. Thus, fibroblast strains have donor-specific antioxidant defense, telomere shortening rate, and growth potential. We used low-density gene expression array analysis of fibroblast strains with different antioxidant potentials and telomere shortening rates to identify gene products responsible for these differences. Extracellular superoxide dismutase was identified as the strongest candidate, a correlation that was confirmed by Northern blotting. Over-expression of this gene in human fibroblasts with low antioxidant capacity increased total cellular superoxide dismutase activity, decreased the intracellular peroxide content, slowed the telomere shortening rate, and elongated the life span of these cells under normoxia and hyperoxia. These results identify extracellular superoxide dismutase as an important antioxidant gene product in human fibroblasts, confirm the causal role of oxidative stress for telomere shortening, and strongly suggest that the senescence-like arrest under mild oxidative stress is telomere-driven.

Mild oxidative stress is not immediately cytostatic. However, it diminishes the replicative capacity of cells. A good experimental example of mild oxidative stress is chronic hyperoxia, which, depending on the oxygen partial pressure and the antioxidant properties of the cells, increases markers of oxidative stress such as malondialdehydes, protein carbonyls, and intracellular peroxides and can seriously limit replicative capacity (8, 13, 14). On the other hand, culture of cells under chronic hypoxia results in a slower accumulation of stress markers (10) and an increased replicative life span (13, 15, 16), indicating that the standard cell culture condition in itself already poses a mild oxidative stress.

Interestingly, using different pro- and antioxidant treatments, telomere shortening per cell division could be varied by 1–2 orders of magnitude (for review, see Ref. 17). A highly significant variation of telomere shortening rates was already found under constant environmental conditions comparing human fibroblast strains from different donors, and this variation was correlated to differences in antioxidant defense capacity (18). However, such differences could not be explained completely by expression patterns of CuZn superoxide dismutase (CuZn-SOD), manganese superoxide dismutase (Mn-SOD), and catalase (19).

It was further shown that oxidative stress-induced single-strand breaks accumulate preferentially in telomeres (7, 20) and that these breaks lead to faster telomere loss during DNA replication (21), possibly by transiently stalling the replication fork (22). These results suggested that a telomere-specific deficiency of single-strand break repair causes the stress-dependence of telomere shortening. However, the gene products responsible for low telomeric repair efficiency are unknown.

To identify possible candidate genes for antioxidant defense and telomere damage repair, we compared the expression patterns of stress-related genes in seven human fibroblast strains with different antioxidant capacity and different telomere shortening rates under conditions of both normoxia and hyperoxia. Of 234 genes, 11 showed a variation in the expression pattern, which correlated to antioxidant capacity and/or te-
EC-SOD Activity Slows Telomere Shortening

**Table I**

<table>
<thead>
<tr>
<th>Gene</th>
<th>5'-Forward primer-3'</th>
<th>5'-Reverse primer-3'</th>
<th>Product size</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mn-SOD</td>
<td>TAC GTG AAC AAC CTG AAC GTC ACC GAG G</td>
<td>GGT AGT AAG CGT GCT CCC ACA CATCAA TCC</td>
<td>400</td>
</tr>
<tr>
<td>EC-SOD I</td>
<td>TTC ATT TGT GAC GAA CCC CGC TCA C</td>
<td>CAA ACA TTC CCC CAA AGG AGG AGC TCT CAG</td>
<td>330</td>
</tr>
<tr>
<td>EC-SOD II</td>
<td>TGG ATC CGA GAC ATG TAC GCC AGG</td>
<td>TCC AGC AGA GGC GAA GGT GAG ACC</td>
<td>738</td>
</tr>
<tr>
<td>GADD153</td>
<td>CTC AGC ATG CCT TGC GAA GAA CC</td>
<td>TGA TTT TGC CCA CAA AGC CTT CC</td>
<td>443</td>
</tr>
<tr>
<td>MTH1</td>
<td>GCC TTT GGG GGC AAA GTG CAA G</td>
<td>CAA TGC CCC CAG GTG AGG ATG G</td>
<td>448</td>
</tr>
<tr>
<td>TOP1</td>
<td>TGC TGA AGA GAC GAA TCA TCG CCC AGG</td>
<td>GAA TGG TAC CCT GAT TGG CAC CG</td>
<td>771</td>
</tr>
<tr>
<td>β-Actin</td>
<td>ATC TGG CAC ACC TTC TAC ATT GAG CTG CG</td>
<td>CGT CAT ACT CCT GCT TGC TGA TCC ACA TCT GC</td>
<td>638</td>
</tr>
</tbody>
</table>

**EXPERIMENTAL PROCEDURES**

**Cell Culture**—Human embryonic lung fibroblasts MRC-5 and WI-38 were obtained from ECACC and ATCC, respectively. Human foreskin fibroblasts (IB strain) originated from the laboratory of J. B. Smith (Houston, TX). All other strains were dermal fibroblasts from adult donors. All strains were extensively characterized in our laboratory in terms of oxidative stress markers, growth potential and telomere length maintenance (12, 18, 23). Cells were grown in Dulbecco’s modified Eagle’s medium plus 10% fetal calf serum in parallel under either normoxia (control) or 40% normobaric hyperoxia using a three-gas cell culture incubator (Zapf Instruments, Sarstedt, Germany). An EC-SOD cDNA clone (24) was obtained from S. L. Marklund (Umea, Sweden). Full-length cDNA was cloned into pBabe-puro and verified by sequencing. MRC-5 fibroblasts at a population doubling (PD) level of 25–30 were transduced with either empty pBabe-puro or pBabe-EC-SOD retrovirus.

**Antioxidative Defense Capacity**—Fibroblasts were grown for at least 1 week in parallel under either normoxia or hyperoxia and then stained with dihydrocholorofluorescin diacetate as described (12). The fluorescence intensity of the intracellularly trapped dihydrocholorofluorescin (DCF) indicates the amount of intracellular peroxides and other oxidizing species. The DCF fluorescence in 2 × 10^5 cells was measured in a flow cytometer (PARTEC, Munster, Germany) using blue excitation and green emission wavelengths. The steady-state antioxidative capacity (AOC) was defined as the ratio DCF(normoxia)/DCF(hyperoxia), resulting in an AOC of 1 for cell strains that can completely counteract the rise in intracellular peroxides under chronic hyperoxic stress.

**Expression Arrays**—Atlas Human Stress Arrays (Clontech) were used according to the manufacturer’s manual. Total RNA from 1–2 × 10^7 cells in logarithmic growth phase was DNase-treated, and poly(A) RNA was enriched by hybridization to biotinylated oligo(dT). Probes were labeled with 50 μCi of [α-32P]dATP during reverse transcription. cDNA was purified by column chromatography and hybridized to the arrays. Quantification was performed using ImageQuantNT software (Amersham Biosciences). The threshold for a hybridization signal was defined as at least two times background variation (determined as the average of all absolute background measurements). Signals lower than this threshold were excluded from the analysis. β-Actin was used as the reference signal. Reproducibility was demonstrated in two strains with independent RNA preparations. For each strain, processing and arraying of normoxia and hyperoxia samples were done in parallel. Threshold values were set at 2-fold for the comparison of expression data between normoxia and hyperoxia within the same strain, and at 3-fold for comparison of expression values between cell strains.

**RT-PCR and Northern Blotting**—RNA was prepared either as described above or using the RNeasy Total RNA kit (Qiagen). Northern blots were probed for Mn-SOD, EC-SOD, GADD153, and β-actin as control. Probes were generated by RT-PCR or by PCR using the EC-SOD cDNA (24) as template with the primers given in Table I. Fragments were gel-purified, tested by restriction analysis, and random prime labeled with [32P]dCTP. ImageQuantNT software (Amersham Biosciences) was used for quantitation.

**Genes for MTH1 and TOP1** were expressed at low level and thus were analyzed by RT-PCR (SuperScript One-Step RT-PCR, Invitrogen). β-Actin was included as the reference control in each sample. Primers are given in Table I. PCR reactions were run for 30, 35, and 38 cycles to control the linearity of amplification. The expression of the EC-SOD transgene was also controlled by RT-PCR. RNA was reverse transcribed using random priming with Truescript RT (Hybaid-AGS), and cDNA equivalent to 50 ng of total RNA was used for a PCR reaction of 30 cycles at 58 °C. Two different non-overlapping primer pairs were used (see Table I). The 788-bp PCR band was cloned, and its correspondence to the cDNA sequence of EC-SOD was confirmed by sequencing.

**SOD Activity**—2 × 10^6 cells were carefully washed and homogenized in 60 μl of phosphate buffer (25). After centrifugation the total SOD activity in the supernatant was measured using the Bioxytech SOD-525 Kit (OXIS Health Products).

**Telomere Length**—Cells at subconfluency were embedded in 0.65% low-melting agarose plugs at a density of 10^7 cells/ml before treatment with proteinase K (7). DNA was digested completely by HinfI (60 units/plug, Roche Molecular Biochemicals) at 37 °C. Plugs were analyzed in a 1% agarose gel by pulsed field gel electrophoresis (BioRad). Gels were blotted to Hybond N+ membranes (Amersham Biosciences) and hybridized with the telomeric probe (TTAGGG)n directly conjugated to alkaline phosphatase (Promega, Madison, WI). A chemiluminescence signal was recorded on film within the linear range and analyzed in an imaging densitometer (BioRad). The average telomere length was calculated as the weighted mean of the optical density as...
described previously (20). The telomere shortening rate per PD was calculated as the slope of the linear regression. To test for nonspecific degradation of DNA, blots were rehybridized with the minisatellite probe (CAC)$_8$, which was end-labeled with $[^{32P}]$ATP (7).

RESULTS

We had shown previously that both the antioxidative capacity (measured as the ability of cells to maintain their peroxidative potential under increased oxygen partial pressure) and the telomere shortening rate varied widely among fibroblast strains derived from different donors (18). Interestingly, both parameters were significantly correlated, suggesting that oxidant stress might be a major cause of telomere shortening in agreement with earlier data (for review, see Ref. 17). To identify genes that might be responsible for the observed differences, we selected seven fibroblast strains, representing the extremes in terms of telomere shortening rate and antioxidative capacity, for array analysis (Fig. 1). Four of the selected strains (BJ, P30, P35, P36) showed low telomere shortening rates and either high (BJ, P36) or moderate (P30, P35) antioxidative capacity, whereas two others (MRC-5, WI-38) displayed a large increase in DCF fluorescence under hyperoxia and high telomere shortening rates. One strain (P13) showed a high telomere shortening rate and a moderate antioxidative capacity.

In these seven cell strains, we measured the expression of 234 genes involved in stress response under both normoxia and hyperoxia using low-density arrays. Of these, 129 genes did not display any significant differences between strains or under increased oxidative stress using the significance levels as described under “Experimental Procedures.” Differences in the expression levels in most of the remaining 105 genes were clearly not correlated to either the telomere shortening rate or AOC. Only a few genes showed an expression pattern that might suggest correlation to cellular peroxide levels or telomere shortening rates. Among these were none of the kinases, cell cycle-related genes, or structural proteins included in the arrays. Those genes with potentially related patterns fall into four groups, namely drug-metabolizing enzymes, heat shock proteins, DNA modification and repair enzymes, and antioxidant enzymes (Fig. 2).

Among the drug-metabolizing enzymes (Fig. 2A), nicotinamide-$n$-methyltransferase (NNMT) was expressed at a significantly lower level in strains with low antioxidative capacity. By catalyzing NAD methylation, NNMT might be involved in free radical defense. On the other hand, glutathione $S$-transferase M1 (GST M1) tended to be higher in MRC-5 and WI-38. GST M1 transfers reduced glutathione to a wide variety of exogenous and endogenous substances to detoxify them.

Three heat shock proteins, HSPB2, HSP90, and HSP47, showed lower mRNA levels in those fibroblast strains with low antioxidative capacity. By catalyzing NAD methylation, NNMT might be involved in free radical defense. On the other hand, glutathione S-transferase M1 (GST M1) tended to be higher in MRC-5 and WI-38. GST M1 transfers reduced glutathione to a wide variety of exogenous and endogenous substances to detoxify them.

Among the DNA modification and repair genes (Fig. 2C), four of them showed expression patterns in the arrays that appeared related to antioxidant capacity and telomere shortening rate (Fig. 2C).
EC-SOD Activity Slows Telomere Shortening

Fig. 3. Confirmation of gene expression patterns. A, semiquantitative RT-PCR for TOP1 (lower band) and β-actin (upper band) in cell strains BJ, P96, P35, and P13. PCR cycle numbers are given above the lanes. M, 100-bp marker (from 600 to 1000 bp). B, correlation of TOP1 expression (signal intensity ratio TOP1/β-actin, 33 cycles) with telomere shortening rates in seven fibroblast strains. The correlation is significant with p = 0.0366. C, Northern blots for EC-SOD and β-actin in 10 fibroblast strains under normoxia (N) and hyperoxia (O).

However, such a correlation remains to be evaluated in the case of replication factor A 2 (RPA2), whereas Northern blotting and semiquantitative RT-PCR failed to confirm a significant correlation of either MTH1 or GADD153 expression with telomere shortening rate (data not shown).

Topoisomerase 1 (TOP1) mRNA expression was at the limit of significance in the array data (Fig. 2C). However, topoisomerases have been implicated in telomere stability in both yeast (26) and human cells (27). Thus, the expression pattern warranted further examination by RT-PCR. Results from seven fibroblast strains showed a significant inverse correlation between TOP1 levels and telomere shortening rates (Fig. 3, A and B).

The mRNA levels for the antioxidant enzymes CuZn-SOD, glutathione peroxidase, heme oxygenase 1, and catalase were generally low and were hardly different between strains and between normoxia and hyperoxia (Fig. 2D). This result is not unexpected in the case of catalase, which is known to be post-transcriptionally regulated. In fact, Western blotting showed significantly higher catalase immunoreactivity in BJ as compared with MRC-5 or WI-38 (not shown). The array analysis indicated a higher Mn-SOD content in BJ fibroblasts as compared with all other strains (Fig. 2D), a result that has been confirmed by Northern blotting (not shown). However, the most striking result was the considerable difference in EC-SOD mRNA levels between cells with low and high peroxide contents (Fig. 2D). Moreover, hyperoxia resulted in a compensatory up-regulation of this mRNA in the two strains with the best antioxidant capacity (BJ and P36).

These results were confirmed by Northern blot (Fig. 3C). The correlation between array and Northern blot results for EC-SOD was significant (p = 0.02, hyperoxia). To establish the relevance of EC-SOD expression more firmly, additional fibroblast strains (including those shown in Fig. 1), which had previously been characterized in terms of DCF fluorescence and telomere shortening rate (18), were included in the analysis. Up-regulation of EC-SOD by hyperoxia was observed in a number of cell strains, especially those with high antioxidant capacity (Fig. 3C). In one strain (F30), up-regulation by hyperoxia was seen by Northern, but not in the arrays, possibly because of a higher noise level in the arrays. For 13 of 18 examined fibroblast strains grown under normoxia, EC-SOD expression correlated strongly with cellular antioxidative capacity (Fig. 4A). Essentially the same result was found for the correlation between EC-SOD expression under hyperoxia and antioxidant capacity (Fig. 4B), although only 11 strains were examined there. The two strains with high EC-SOD expression but comparatively low antioxidant capacity (Fig. 4A) displayed conspicuous deregulation of the catalase protein content (data not shown). We have not yet identified a plausible explanation for the deviation from the common trend toward high antioxidant capacity in three strains.

EC-SOD expression is not only a strong statistical predictor of antioxidant defense in most fibroblast strains; it is also significantly inversely correlated to the rate of telomere shortening both under normoxia (Fig. 4C) and under hyperoxia (Fig. 4D). Interestingly, even strains that do not fit the general trend, in Fig. 4, A and B, did not obviously deviate from the correlations shown in Fig. 4, C and D, which might indicate that a moderately good antioxidant capacity suffices to maintain a low telomere shortening rate. Taken together, these correlative data point to EC-SOD as the strongest candidate for maintaining low levels of oxidative stress in human fibroblasts under hyperoxia and low rates of telomere shortening.

To discover whether cellular SOD activity is dependent on the level of transcription of EC-SOD mRNA, we measured total SOD activity in cell extracts from the most extreme cell strains, BJ and MRC-5. SOD activity in BJ cells was significantly higher than that in MRC-5 (Fig. 5). Retroviral transduction of an EC-SOD cDNA (24) resulted in an at least a 10-fold over-expression of EC-SOD mRNA in MRC-5 fibroblasts as judged by RT-PCR (not shown). This increased the total cellular SOD...
EC-SOD Activity Slows Telomere Shortening

Fig. 5. Total SOD activity in cell extracts (units/million cells) in BJ fibroblasts (BJ), parental MRC-5 (Mpar), MRC-5 transduced with empty virus (MpB), and MRC-5 over-expressing EC-SOD (MSOD). Data are mean ± S.E. from two to eight experiments. The SOD activity in BJ and EC-SOD-over-expressing MRC-5 cells is significantly higher than in parental or pBabe-transfected MRC-5 fibroblasts (ANOVA, p < 0.05).

Fig. 6. EC-SOD expression diminishes oxidative stress and increases growth capacity. A, median DCF fluorescence in arbitrary units (a.u.) of parental MRC-5 (C), MRC-5 transduced with empty virus (pB), and MRC-5 over-expressing EC-SOD (SOD) under normoxia (open bars) and 40% hyperoxia (filled bars). Data are mean ± S.E. from triplicate measurements. DCF fluorescence intensities in EC-SOD-expressing cells are significantly lower than in either parental cells or vector controls under both normoxia and hyperoxia (ANOVA, p < 0.05). B, growth capacity (population doublings achieved) under normoxia (open bars, left scale) and under 40% hyperoxia (filled bars, right scale). Data are mean ± S.E. from five independent experiments (hyperoxia) and from one set of parallel experiments (normoxia). The growth capacity of EC-SOD-expressing cells under hyperoxia is significantly higher than that of either parental cells or vector controls (ANOVA, p < 0.05). The difference between control and vector-transduced cells is not significant.

Over-expression of EC-SOD markedly decreased the concentration of intracellular peroxides as measured by cellular median DCF fluorescence intensity, both under normoxia and under hyperoxia (Fig. 6A). In accordance with their lower antioxidant capacity, MRC-5 fibroblasts achieve significantly less population doublings than BJ fibroblasts, both under normoxia and, even more so, under hyperoxia (7, 12). The replicative life span of MRC-5 fibroblasts under normoxia increased by about 25% after transfection with EC-SOD as compared with parallel vector only-transfected controls (Fig. 6B). This is comparable with the magnitude of effect of typical antioxidants and free radical scavengers (7, 8, 16). When parental MRC-5 and vector-transfected cells were transferred to hyperoxia at a PD of about 30, they became senescent after about 1 PD in accordance with earlier results (7, 8). EC-SOD over-expression sustained growth of MRC-5 cells under hyperoxia more than twice as long (Fig. 6B).

To examine whether EC-SOD expression would influence the telomere shortening rate of MRC-5 cells, we measured telomere lengths in both vector-transfected and EC-SOD-over-expressing MRC-5 fibroblasts. Cells transduced with empty virus showed an even faster rate of telomere shortening under hyperoxia than parental ones. This was not because of unspecific DNA degradation, as shown by rehybridization of the blots with an interstitial minisatellite probe (Fig. 7A). Mitochondrial uncoupling results in an even more exaggerated telomere shortening within a single cell cycle in mouse zygotes (28). Over-expression of EC-SOD resulted in a significantly lower shortening rate under both normoxia and hyperoxia (Fig. 7). Telomere shortening in EC-SOD-over-expressing fibroblasts under hyperoxia was even slower than in control cells under normoxia. Thus, increased expression of EC-SOD both slows down telomere shortening and postpones replicative senes-
cence in MRC-5 fibroblasts under normoxic and hyperoxic culture conditions.

**DISCUSSION**

We used low-density arrays of stress-related genes to identify possible candidates involved in the control of telomere shortening rate and antioxidant capacity in human fibroblasts under mild oxidative stress. This resulted in the identification of only one of the DNA modification and repair enzymes; high TOP1 expression was correlated significantly to low telomere shortening rate. A number of data link the action of topoisomerases to telomeres. Loss of topoisomerase III leads to telomere shortening in yeast (26). Topoisomerase II was shown to cleave telomeres preferentially (29). Topoisomerases interact with members of the RecQ helicase family (30), which are known to bind telomeres. Telomerase activity modifies the sensitivity of human cells to topoisomerase inhibitors (27). TOP1 is involved in the resolution and repair of single-strand breaks in telomeres and are a major determinant of the telomere shortening rate (7, 21). It will be interesting, therefore, to examine the possible role of TOP1 in telomere repair and length maintenance more closely.

Our array studies revealed that the expression of only one superoxide dismutase, EC-SOD, correlated strongly with antioxidant defense capacity in human fibroblast strains. Steady-state levels of EC-SOD mRNA in antioxidatively competent strains were between 5 and 20 times higher than levels of CuZn-SOD. They were also 5–20 times higher than EC-SOD levels in incompetent strains. This is in agreement with earlier data showing up to 100-fold variation in EC-SOD expression between human fibroblast strains (25). EC-SOD expression was found to be inducible by mild chronic hyperoxia in some (generally antioxidatively competent) strains but not in others. Such noninducibility has been reported previously (31). Strains with high expression of EC-SOD (either endogenously or from a transgene) showed high total SOD activity in cellular lysates, suggesting that EC-SOD contributes significantly to the cellular superoxide dismutase activity. It has previously been shown that between 5 and 25% of EC-SOD protein is not secreted but is retained in the cell lysates (25). This retained fraction is not bound to extracellular heparin sulfate proteoglycan, but little more is known about its distribution.

The two most obvious locations for cell-bound EC-SOD protein are at the plasma membrane and within the endolysosomal system. In both locations, the enzyme might contribute significantly to cellular antioxidant defenses; NADH oxidation by the plasma membrane oxido-reductase system has recently been suggested as a likely source of superoxide (32). SOD activity at the plasma membrane might thus block the generation of reactive oxygen species at source. Alternatively, lysosomes are a major site for cellular iron storage and are thus highly susceptible to damage by hydrogen peroxide (33). The presence of EC-SOD in the endolysosomal system could inhibit superoxide-dependent recycling of catalytically active iron.

In line with these hypotheses, over-expression of EC-SOD was found to decrease the amount of cellular peroxides and to slow down telomere shortening, under both standard conditions and increased oxidative load, i.e. hyperoxia. This result confirms that telomere shortening under conditions of low to mild oxidative stress is to a large extent caused by telomeric DNA damage (7, 17).

The senescence-like growth arrest of human fibroblasts exposed to mild oxidative stress is undistinguishable from “Hayflick senescence” under standard culture conditions, in terms of telomere length (8, 23) and phenotype (34, 35). The fact that over-expression of an antioxidant, EC-SOD, slows down telomere shortening and extends the replicative life span under both normoxia and hyperoxia in parallel strongly suggests that senescence under such mild oxidative stress conditions is telomere-driven.

In mice, EC-SOD over-expression or knockout phenotypes are comparatively mild. However, EC-SOD knockout mice are more sensitive to hyperoxia (36), and this increased sensitivity does not seem to be due to modification of the inflammatory response in the lungs (37). More interestingly, EC-SOD plays an important role in brain function, as both young EC-SOD-over-expressing and knockout mice are impaired in spatial learning and memory, which has been attributed to a deregulation of nitric oxide catabolism (38, 39). Conversely, knockout mice are more sensitive to focal cerebral ischemia (40), and EC-SOD over-expression increases the resistance of mice to global ischemia (41). Furthermore, improved learning and memory performance in aged mice over-expressing EC-SOD has been demonstrated recently (42). Together, these data suggest a protective role of EC-SOD in the brain, possibly related to its antioxidant function rather than to its role in nitric oxide turnover. In humans, we found short telomeres in patients with stroke-related (vascular) dementia. We assumed that short telomeres might indicate low antioxidant capacity and thus higher risk of stroke-induced brain damage in vivo (18). Data demonstrating a protective role for EC-SOD against ischamic brain damage (40, 41) and age-related deterioration (42) together with the demonstration of a role for EC-SOD in telomere maintenance in human cells (this study) are in agreement with this interpretation.

Acknowledgments—We thank Stefan Marklund (Umea, Sweden) for the gift of EC-SOD cDNA and for helpful discussions. We thank B. Keys for expert technical assistance and C. Martin-Ruiz for critical reading the manuscript.

**REFERENCES**

EC-SOD Activity Slows Telomere Shortening

MOLECULAR BASIS OF CELL AND DEVELOPMENTAL BIOLOGY: 
Extracellular Superoxide Dismutase Is a Major Antioxidant in Human Fibroblasts and Slows Telomere Shortening

Violeta Serra, Thomas von Zglinicki, Mario Lorenz and Gabriele Saretzki

doi: 10.1074/jbc.M207939200 originally published online December 9, 2002

Access the most updated version of this article at doi: 10.1074/jbc.M207939200

Find articles, minireviews, Reflections and Classics on similar topics on the JBC Affinity Sites.

Alerts:
- When this article is cited
- When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

This article cites 41 references, 14 of which can be accessed free at http://www.jbc.org/content/278/9/6824.full.html#ref-list-1